Synthesis and Functionalization of Double Picket Fence Porphyrins Starting from 4.6-Disubstituted **Pyrimidine-5-carbaldehydes**

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Introduction

The ortho-substituted and ortho.ortho'-disubstituted tetraarvlporphyrins have been subject of intensive research. In general, sterically encumbered porphyrins have served as useful models for some aspects of the chemistry of hemoproteins.¹ The "picket fence",² "double picket fence", and "capped"³ porphyrins have been studied as models for hemoproteins and as catalysts. Ortho, ortho'-substituted tetraarylporphyrins or "double picket fence" porphyrins have been used extensively in secondgeneration oxidation catalysts.⁴ Membranes of picketfence cobalt porphyrins have been studied as efficient oxygen carriers.⁵ Ferric porphyrins bearing chiral binaphthalene moieties on both faces were used in enantioselective oxidation of sulfides and in asymmetric epoxidations.⁶ These "twin coronet" porphyrins have been further modified to mimic the active center of heme enzymes such as cytochrome P-450.7 Functionalization can be achieved with electrophiles on 5,10,15,20-tetrakis-

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(2,6-dihydroxyphenyl)porphyrin⁸ and the analogous aminosubstituted tetraarylporphyrins.⁹ However, in general the synthesis of these porphyrins is not straightforward. For instance the condensation of 2,6-dinitrobenzaldehyde with pyrrole has been reported to give the octanitro precursor in only in 0.75% yield in the presence of TFA¹⁰ while the use of BF₃·Et₂O as catalyst gave 2-13% yield.¹¹ Better yields could be achieved using 2,6-dinitro-4-tertbutyl-benzaldehyde, where the tert-butyl group increased the solubility of the intermediates.⁴ However, the latter aldehyde could only be obtained involving three steps in an overall yield of 41% starting from 4-tert-butyltoluene. The reduction toward the ortho, ortho'-amino-substituted porphyrins could be achieved in 60% yield. Condensation of 2,6-dimethoxybenzaldehyde with pyrrole yielded the octamethoxy precursor (12%).¹² This porphyrin was then demethylated using BBr₃ giving an overall yield of 9%.^{8c} Nucleophilic substitution of the chlorine atoms of the readily available 5,10,15,20-tetrakis(2',6'-dichlorophenyl)porphyrin¹³ is apparently not feasable. However, functionalization by electrophilic substitution at the meso aryl position could be achieved using 3- or 4-nitro-substituted 2,6-dichlorobenzaldehyde. The nitro group was reduced to amine which could then react further with electrophiles. It should be noted that tailed porphyrins bearing amido bonds are less stable than those having an ether linkage.14

Results and Discussion

In this paper we describe the synthesis of several double picket fence porphyrins using 4,6-disubstituted pyrimidine-5-carbaldehydes. The 4,6-dichloro-pyrimidine-5-carbaldehyde 1 can be considered as a heterocyclic analogue of 2,6-dichlorobenzaldehyde. The chlorine functionalities on the pyrimidine ring are highly activated toward nucleophilic substitution, allowing us to introduce substituents at the aldehyde as well as the porphyrin stage.15

The aldehyde 1 was prepared using the reported Vilsmeier conditions (DMF/POCl₃) for chloroformylation of 4,6-dihydroxypyrimidine.¹⁶ Functionalization of the two chlorine atoms of aldehyde 1 was carried out with several substituted phenolates. Typically, the nucleo-

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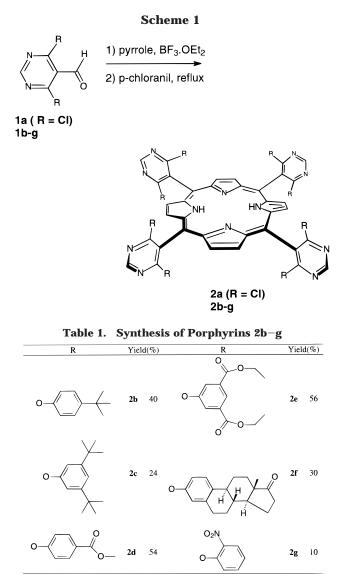
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philic substitution of **1a** was carried out using 2.2 equiv of phenol and an excess K_2CO_3 (5-fold) under reflux in THF for 5 h. This yielded the 4,6-disubstituted pyrimidine-5-carbaldehydes **1b**-g in excellent yields.

These aldehydes were reacted with pyrrole under Lindsey conditions (Scheme 1).¹⁷ The porphyrins 2b-ewere isolated in yields that ranged from good to excellent (24–59%) (Table 1). On introduction of a nitro group in *ortho*-position of phenol (aldehyde 1g) the yield of the corresponding porphyrin decreased noticeably (6%). This relatively low yield is not only a consequence of the highly sterically hindered aldehyde but also of the low solubility of the corresponding intermediates in the reaction mixture (solvent: dichloromethane).

Working under more diluted conditions $(3.4 \times 10^{-3} \text{ M})$ instead of $3.4 \times 10^{-2} \text{ M}$) gave an increased yield of 10%. We were not able to isolate **2a**, the heterocyclic analogue of 5,10,15,20-tetrakis(2',6'-dichlorophenyl)porphyrin using either Adler–Longo (refluxing propionic acid) or Lindsey conditions. Again, this may be a result of low solubility of the intermediates. On the other hand, the isolated yields after chromatographic separation and

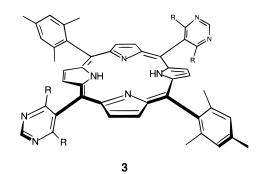
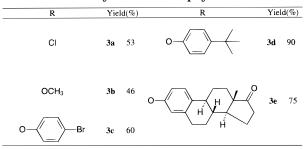


Figure 1.

Table 2. Synthesis of Porphyrins 3a-e



crystallization of double picket fence porphyrins **2d** (54%) and **2e** (59%) are among the highest ever obtained for a Rothemund porphyrin synthesis.

It might still be more effective to introduce the substituents directly at the porphyrin stage by substitution of the chlorine functions of the meso-(2,6-dichloropyrimidyl) substituents. Unfortunately, octachloro porphyrin 2a is not available, as mentioned above. Mixed Rothemund condensations, as reported by us in a previous communication, give statistical mixtures of several pyrimidyl porphyrins which are difficult to separate. Therefore, a 5,15-bis(pyrimidyl)porphyrin **3a** was prepared by a Mc-Donald condensation of mesityldipyrromethane¹⁸ with aldehyde 2a. In previous work we reported that the substitution of **3a** with methoxide gave an inseparable mixture of disubstituted atropoisomers. This reaction was carried out in THF at reflux for 72 h in the presence of K₂CO₃. We now carried out the substitution reaction in DMF using sodium methoxide. We were able to obtain the tetramethoxy-substituted porphyrin 3b. Reaction of porphyrin **3a** with several phenols (see Figure 1) in the presence of potassium carbonate at 60°C in DMF resulted in the formation of four-picket porphyrins 3d (60%), 3c (90%), 3e (75%) in high yields (Table 2). In case of expensive phenol starting materials (e.g., estrone), this method is clearly superior.

Conclusions

Double picket fence porphyrins which were highly sterically crowded could be prepared in high yields either from pyrimidine or porphyrin starting materials. Substituents are easily introduced on both faces of the porphyrins, which leads to three-dimensionally structured macromolecules. They have been developed as model compounds for oxygen-binding and oxygen-activat-

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ing heme proteins,¹⁹ octopus porphyrins²⁰ that form monolayer assemblies, functional dendrimers²¹ and donor– acceptor systems²² of potential applications. Enantioselective catalysts can be designed by the introduction of chiral substituents close to the catalytic center of metalloporphyrins.^{6,23} The three-dimensional architecture of such molecules would also make them valuable as host molecules for stereoselective molecular recognition.²⁴

Experimental Section

General. Mass spectra were obtained on a Hewlett-Packard MS instrument nE 5989A in EI (250 °C), on a Micromass Quatro II in ESI (infusion of 50 il MeOH/CH₂Cl₂–NH₄OAc (0.1M in MeOH) with a Harvard pump, model 11), and on a Micromass Quatro II in APCI (250 μ L of MeOH/CH₂Cl₂ using a Hewlett-Packard HP 1100 binary pump and infusion of 20–30 μ L of MeOH/CH₂Cl₂–NH₄OAc (0.1M in MeOH) with a Harvard pump model 11).

General Procedure for the Synthesis of 4,6-Disubstituted Pyrimidine-5-carbaldehydes. To a well-stirred suspension of anhydrous potassium carbonate (1.52 g, 11 mmol) in THF (100 mL) the substituted phenol (4.4 mmol) was added, and the mixture was stirred with refluxing. After 0.5 h, 4,6-dichloropyrimidine-5-carbaldehyde **1a** (0.352 g, 2 mmol) was added, and the stirring and refluxing was continued for 5 h. The reaction mixture was cooled, and solvent was removed under vacuum. The residue was taken in dichloromethane (100 mL) and washed with water (3 × 30 mL). The organic layer was dried over magnesium sulfate and evaporated to give the crude product, which precipitated on addition of methanol. The resulting suspension was filtered to give the substituted pyrimidinecarbaldehydes **1b**–**g** as colorless solids. They were further purified by recrystallization (methylene chloride/hexane, 1:1).

4,6-Bis[**4**-(*tert*-butyl)**phenoxy**]**pyrimidine**-**5**-carbaldehyde (1b). Reaction of 4,6-dichloropyrimidine-**5**-carbaldehyde (0.352 g, 2 mmol) and 4-*tert*-butylphenol (0.660 g, 4.4 mmol) gave **1b** (0.735 g, 91%): mp 160–161 °C; ¹H NMR (400 MHz, CDCl₃) δ 1.36 (s, 18H), 7.12 (d, J = 8.7 Hz, 4H), 7.47 (d, J = 8.7 Hz, 4H), 8.45 (s, 1H), 10.66 (s, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 31.42, 120.95, 126.65, 149.03, 149.77, 160.97, 170.23, 185.73; IR-(KBr), v = 2960, 2866, 1701, 1552, 1506, 1420 cm⁻¹; EI-MS 404 (M⁺); Anal. Calcd for C₂₅H₂₈N₂O₃: C, 74.23; H, 6.98; N, 6.93. Found: C, 74.22; H, 7.06; N, 6.85.

4,6-Bis[3,5-di(*tert*-butyl)phenoxy]pyrimidine-5-carbaldehyde (1c). Reaction of 4,6-dichloropyrimidine-5-carbaldehyde

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4,6-Bis(4-methoxycarbonylphenoxy)pyrimidine-5-carbaldehyde (1d). Reaction of 4,6-dichloropyrimidine-5-carbaldehyde (0.352 g, 2 mmol) and methyl 4-hydroxybenzoate (0.669 g, 4.4 mmol) gave **1d** (0.718 g, 88%): mp 223–224 °C; ¹H NMR (400 MHz, CDCl₃) δ 3.94 (s, 6H), 7.27 (d, J = 6.8 Hz, 4H), 7.36 (s, 2H), 8.16 (d, J = 6.8 Hz, 4H), 8.38 (s, 1H), 10.65 (s, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 52.25, 103.53, 121.79, 128.26, 131.46, 155.55, 160.67, 166.07, 169.66, 185.02; IR(KBr), 1735, 1702, 1583, 1552, 1441 cm⁻¹; APCI-MS 409 (MH)⁺. Anal. Calcd for C₂₁H₁₆N₂O₇ C, 61.77; H, 3.95; N; 6.86. Found: C, 61.63; H, 3.95; N, 6.70.

4,6-Bis[3,5-bis(ethoxycarbonyl)phenoxy]pyrimidine-5carbaldehyde (1e). Reaction of 4,6-dichloropyrimidine-5-carbaldehyde (0.352 g, 2 mmol) and diethyl isophthalate (1.05 g, 4.4 mmol) gave **1e** (1.115 g, 96%): mp 147–148 °C; ¹H NMR (400 MHz, CDCl₃) δ 1.42 (t, J = 7.2 Hz, 6H), 4.43 (q, J = 7.2Hz, 4H), 8.06 (d, J = 1.48 Hz, 4H), 8.37 (s, 1H), 8.64 (t, J = 1.48Hz, 2H), 10.68 (s, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 14.26, 61.72, 103.43, 127.14, 128.35, 132.72, 151.93, 160.59, 164.73, 169.68, 184.82; IR(KBr) v = 2976, 1740, 1721, 1700, 1553 cm⁻¹; EI-MS 580 (M⁺). Anal. Calcd for C₂₉H₂₈N₂O₁₁ C, 60.00; H, 4.86; N, 4.83. Found: C, 60.01; H, 4.83; N, 4.66.

4,6-Bis[estra-17-oxo-1,3,5(10)-trien-3-oxy]pyrimidine-5-carbaldehyde (1f). Reaction of 4,6-dichloropyrimidine-5-carbaldehyde (0.352 g, 2 mmol) and estrone (1.190 g, 4.4 mmol) gave **1d** (0.953 g, 74%): mp >250 °C; ¹H NMR (400 MHz, CDCl₃) δ 0.92 (s, 6H), 1.45–1.66 (m, 12H), 1.96–2.18 (m, 8H), 7.61 (t, J = 8.0 Hz, 2H), 7.68 (d, J = 8.0 Hz, 2H), 7.90 (t, J = 8.0 Hz, 2H), 8.24 (d, J = 8.0 Hz, 2H), 8.43 (s, 1H), 10.56 (s, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 102.43, 125.46, 125.84, 127.55, 135.82, 141.27, 144.43, 160.46, 168.61, 185.01; IR(KBr), v = 2928, 2858, 1737, 1700, 1552, 1491 cm⁻¹; CI-MS 645 (MH)⁺. Anal. Calcd for C₄₁H₄N₂O₅ C, 76.37; H, 6.88; N, 4.34. Found: C, 76.17; H, 6.81, N, 4.15.

4,6-Bis(2-nitrophenoxy)pyrimidine-5-carbaldehyde (1g). Reaction of 4,6-dichloropyrimidine-5-carbaldehyde (0.352 g, 2 mmol) and 2-nitrophenol (0.612 g, 4.4 mmol) gave **1g** (0.642 g, 84%): ¹H NMR (400 MHz, CDCl₃) δ 7.61 (t, J = 8.0 Hz, 2H), 7.68 (d, J = 8.0 Hz, 2H), 7.90 (t, J = 8.0 Hz, 2H), 8.24 (d, J = 8.0 Hz, 2H), 8.43 (s, 1H), 10.56 (s, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 102.43, 125.46, 125.84, 127.55, 135.82, 141.27, 144.43, 160.46, 168.61, 185.01; IR(KBr), v = 1703, 1568, 1530, 1506, 1343 cm⁻¹; CI-MS 383 (MH)⁺. Anal. Calcd for C₁₇H₁₀N₄O₇ C, 53.41; H, 2.64; N, 14.66. Found: C, 53.48; H, 2.65; N, 14.60.

General Procedure for the Synthesis of Porphyrins 2b– g. A 0.05 M solution of pyrimidinecarbaldehyde **1b–g** and pyrrole (1 equiv) in dichloromethane was deoxygenated by purging with argon for 15 min. Boron trifluoride etherate (0.1 equiv) was added by syringe, and the solution was stirred under argon and protected from light for 2 h at 20 °C. *p*-Chloranil (1 equiv) was added, and the mixture was refluxed for 1 h. The solvent was removed, and the residue was chromatographed (silica gel; CH₂Cl₂). The obtained product was redissolved in dichloromethane and precipitated with MeOH yielding **2b–g** as purple solids.

5,10,15,20-Tetrakis[**4,6-bis**[**4**-(*tert*-**butyl**)**phenoxy**]**pyrimidin-5-yl**]**porphyrin (2b)** was obtained from aldehyde **1b** (0.650 g, 1.61 mmol) following the general procedure (0.300 g, 40%): ¹H NMR (400 MHz, CDCl₃) δ –2.45 (s, 2H), 1.12 (72H, s), 6.93 (d, J = 8.8 Hz, 16H), 7.16 (d, J = 8.8 Hz, 16H), 8.91 (s, 4H), 9.15 (s, 8H); UV–vis (CH₂Cl₂): λ_{max} (log ϵ) 422.1 (5.5978), 516.4 (4.2845), 549.3 (3.7736), 592.0 (3.8478), 646.9 (3.3078); APCI-MS 1807.9 (MH⁺); Anal. Calcd for C₁₁₆H₁₁₈N₁₂O₈ C, 77.05; H, 6.58; N, 9.29. Found: C, 76.98; H, 6.65; N, 9.33.

5,10,15,20-Tetrakis[**4,6-bis**]**3,5-bis**(*tert*-**butyl**)**phenoxy**]-**pyrimidin-5-yl**]**porphyrin (2c)** was obtained from aldehyde **1c** (0.500 g, 0.97 mmol) following the general procedure (0.135 g, 24%): ¹H NMR (400 MHz, CDCl₃) δ –2.45 (s, 2H), 1.12 (s, 144H), 6.80 (d, J = 1.6 Hz, 16H), 7.07 (d, J = 1.6 Hz, 16H), 8.88

(s, 4H), 9.21 (s, 8H); UV–vis (CH₂Cl₂): λ_{max} (log ϵ) 422.1 (5.5978), 516.4 (4.2845), 549.3 (3.7736), 592.0 (3.8478), 646.9 (3.3078); APCI-MS 2056.4 (MH⁺); Anal. Calcd for C₁₄₈H₁₈₂N₁₂O₈ C, 78.76; H, 8.13; N, 7.45. Found: C, 78.60; H, 8.00; N, 7.43.

5,10,15,20-Tetrakis[4,6-bis(4-methoxycarbonylphenoxy)pyrimidin-5-yl]porphyrin (2d) was obtained from aldehyde 1d (0.500 g, 1.22 mmol) following the general procedure (0.303 g, 54%): ¹H NMR (400 MHz, CDCl₃) δ –2.37 (s, 2H), 3.83 (s, 24H), 7.04 (d, J = 8.8 Hz, 16H), 7.89 (d, J = 8.8 Hz, 16H), 8.87 (s, 4H), 9.16 (s, 8H); UV–vis (CH₂Cl₂): λ_{max} (log ϵ) 422.0 (5.6952), 518.1 (4.3272), 547.8 (3.5906), 591.6 (3.7848), 647.03 (2.2552); APCI-MS 1823.5 (MH⁺). Anal. Calcd for C₁₀₀H₇₀N₁₂O₂₄ C, 65.86; H, 3.87; N, 9.22. Found: C, 65.46; H, 3.97; N, 8.97.

5,10,15,20-Tetrakis(4,6-bis(3,5-bis(ethoxycarbonyl)phenoxy)pyrimidin-5-yl)porphyrin (2e) was obtained from aldehyde **1e** (0.500 g, 0.86 mmol) following the general procedure (0.320 g, 59%): ¹H NMR (400 MHz, CDCl₃) δ –2.29 (s, 2H), 1.25 (t, *J* = 7.2 Hz, 48H), 4.24 (q, *J* = 7.2 Hz, 32H), 7.87 (d, *J* = 1.4 Hz, 16H), 8.40 (t, *J* = 1.4 Hz, 8H), 8.84 (s, 4H), 9.28 (s, 8H); UV-vis (CH₂Cl₂): λ_{max} (log ϵ) 421.3 (5.6727), 515.1 (4.3407), 545.9 (3.6300), 590.4 (3.8511), 645.0 (3.0256); ES-MS 2511.8 (MH⁺); Anal. Calcd for C₁₃₂H₁₁₈N₁₂O₄₀ C, 63.10; H, 4.73; N, 6.69. Found: C, 63.02; H, 4.72; N, 6.54.

5,10,15,20-Tetrakis[4,6-bis[estra-17-oxo-1,3,5(10)-trien-3-oxy]-pyrimidin-5-yl]porphyrin (2f) was obtained from aldehyde 1f (0.810 g, 1.26 mmol) following the general procedure (0.260 g, 30%): ¹H NMR (400 MHz, CDCl₃) δ –2.54 (s, 2H), 0.68 (s, 24H), 1.05–1.60 (m, 48H), 1.65–2.45 (m, 56H), 2.50–265 (m, 16H), 6.65 (d, J = 2.3 Hz, 8H), 6.79 (dd, J = 8.5, 2.3 Hz, 8H), 7.03 (d, J = 8.4 Hz, 8H), 8.91 (s, 4H), 9.18 (s, 8H); UV–vis (CH₂-Cl₂): λ_{max} (log ϵ) 422.3 (5.8151), 516.8 (4.5928), 547.7 (4.1209), 593.0 (4.1673), 647.2 (3.8322); APCI-MS 2768.4 (MH⁺). Anal. Calcd for Cl₁₈₀H₁₈₂N₁₂O₁₆ C, 78.06; H, 6.62; N, 6.07. Found: C, 78.05; H, 6.54; N, 6.19.

5,10,15,20-Tetrakis[**4,6-bis**(**2**-nitrophenoxy)**pyrimidin-5yl**]**porphyrin (2g)** was obtained from aldehyde **1g** (0.500 g, 1.31 mmol) following the general procedure (0.035 g, 6.2%). Modification of this procedure using a 5 × 10⁻³ M solution yielded porphyrin **2g** (0.053 g) in 10% yield; ¹H NMR (400 MHz, DMSO) δ – 2.71 (s, 2H), 7.40 (d, J = 5.8 Hz, 16H), 7.77 (t, J = 7.4 Hz, 8H), 8.02 (d, J = 8.0 Hz, 8H), 8.83 (s, 4H), 9.54 (s, 8H); UV–vis (THF): λ_{max} (log ϵ) 419.8 (4.4651), 513.2 (4.2741), 540.0 (4.0840), 590.2 (4.0905); ESI-MS 1718.3 (MH⁺). Anal. Calcd for C₈₄H₄₆N₂₀O₂₄: C, 58.68; H, 2.70; N, 16.29. Found: C, 58.61; H, 2.79; N, 16.00.

5,15-Bis(4,6-dichloropyrimidin-5-yl)-10,20-bis(2,4,6-trimethylphenyl)porphyrin (3a). A solution of 2,4,6-trimethylphenyldipyrromethane (0.500 g, 1.89 mmol) and 4,6-dichloropyrimidine-5-carbaldehyde (0.337 g, 1.89 mmol) in dichloromethane (500 mL) was deoxygenated by purging with argon for 15 min. Boron trifluoride etherate (0.020 g, 0.19 mmol) was added by syringe, and the solution was stirred under argon and protected from light for 1 h at room temperature. p-Chloranil (0.454 g, 1.89 mmol) was added, and the mixture was refluxed for 1 h. The solvent was removed and the residue chromatographed (silica gel, dichloromethane) yielding porphyrin 3 as a purple solid (0.445 g, 53%): ¹H NMR (400 MHz, CDČl₃) δ -2.50 (s, 2H), 1.86 (s, 12H), 2.63 (s, 6H), 7.29 (s, 4H), 8.59 (d, J = 4.8 Hz, 4H), 8.76 (d, J = 4.8 Hz, 4H), 9.26 (s, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 21.45, 21.75, 108.97, 120.00, 127.96, 129.03, 131.88, 134.72, 137.28, 138.30, 139.35, 158.22, 164.06; UV-vis (CH₂Cl₂): λ_{max} $(\log \epsilon)$ 419.4 (5.8111), 514.6 (4.4410), 548.1 (3.8030), 591.2 (3.8255), 647.9 (3.5997); ESI-MS 839.2 (MH⁺). Anal. Calcd for C43H34N14O8: C, 65.72; H, 4.08; N, 13.33. Found: C, 65.81; H, 4.11; N, 13.36.

General Procedure for Synthesis of Porphyrins 3c-e. To a solution of porphyrin **3a** (0.050 g, 0.06 mmol) and substituted phenol (4.4 equiv) in dry DMF (4 mL) was added K_2CO_3 (8.8 equiv). The mixture was stirred for 72 h at 60 °C under argon atmosphere. The solution was allowed to reach room temperature, and dichloromethane (20 mL) was added. The organic layer was washed three times with water (3 \times 20 mL) and dried over MgSO₄. The solvent was removed and the residue purified by column chromatography (silica gel; dichloromethane/hexane 3/1). The obtained product was redissolved in dichloromethane and precipitated with MeOH yielding **3b**-**d** as purple solids.

5,15-Bis[**4,6-di**(**4-bromophenoxy**)**pyrimidin-5-yl**]-**10,20bis**(**2,4,6-trimethylphenyl**]**porphyrin** (**3c**) was obtained from 4-bromophenol following the general procedure (0.074 g, 90%): ¹H NMR (400 MHz, CDCl₃) δ –2.43 (s, 2H), 1.86 (s, 12H), 2.64 (s, 6H), 6.87 (d, J = 8.9 Hz, 8H), 7.31 (s, 4H), 7.32 (d, J = 8.9 Hz, 8H), 8.78 (d, J = 4.7 Hz, 4H), 8.81 (s, 2H), 8.94 (d, J = 4.7 Hz, 4H); UV–vis (CH₂Cl₂): λ_{max} (log ϵ) 419.1 (5.7318), 514.5 (4.3567), 548.1 (3.9324), 591.0 (3.8667), 646.6 (3.6673); APCI-MS 1383.0/1385.0/1387.0/1389.0/1391.0 (MH⁺). Anal. Calcd for C₇₀H₅₀Br₄N₈O₄ C, 60.63; H, 3.63; N, 8.08. Found: C, 60.49; H, 3.74; N, 7.80.

5,15-Bis[4,6-bis[4-(*tert***-butyl)phenoxy]pyrimidin-5-yl]-10,20-bis(2,4,6-trimethylphenyl)porphyrim (3d)** was obtained from 4-*tert*-butylphenol following the general procedure (0.050 g, 60%): ¹H NMR (400 MHz, CDCl₃) δ –2.45 (s, 2H), 1.16 (s, 36H), 1.88 (s, 12H), 2.65 (s, 6H), 6.90 (d, J = 8.8 Hz, 8H), 7.19 (d, J = 8.8 Hz, 8H), 7.31 (s, 4H), 8.76 (d, J = 4.7 Hz, 4H); 8.86 (s, 2H), 8.99 (d, J = 4.7 Hz, 4H); UV-vis (CH₂Cl₂): λ_{max} (log ϵ) 419.2 (5.6847), 514.8 (4.3141), 548.4 (3.8706), 591.2 (3.8047), 646.9 (3.5729); APCI-MS 1295.7 (MH⁺). Anal. Calcd for C₈₆H₈₆N₈O₄: C, 79.72; H, 6.69; N, 8.65. Found: C, 79.60; H, 6.79; N, 8.65.

5,10-Bis[4,6-bis[estra-17-oxo-1,3,5(10)-trien-3-oxy]pyrimidin-5-yl]-10,20-bis(2,4,6-trimethylphenyl)porphyrin (3e) was obtained from estrone following the general procedure (0.080 g, 75%): ¹H NMR (400 MHz, CDCl₃) δ –2.44 (s, 2H), 0.79 (s, 12H), 1.20–1.60 (m, 24H), 1.86 (s, 12H), 1.80–2.20 (m, 20H), 2.22–2.29 (m, 4H), 2.38–2.48 (m, 4H), 2.65 (s, 6H), 2.68–2.76 (m, 8H), 6.67 (d, J = 2.4 Hz, 4H), 6.79 (dd, J = 8.6, 2.4 Hz, 4H), 7.12 (d, J = 8.6 Hz, 4H), 7.31 (s, 4H), 8.77 (d, J = 4.7 Hz, 4H), 8.86 (s, 2H), 8.99 (d, J = 4.7 Hz, 4H); UV–vis (CH₂Cl₂): λ_{max} (log ϵ) 419.5 (5.6662), 515.1 (4.3235), 548.2 (3.8699), 591.4 (3.8123), 647.1 (3.5959); APCI-MS 1777.2 (MH⁺). Anal. Calcd for C1₁₁₈H₁₁₈N₈O₈ C, 79.79; H, 6.70; N, 6.31. Found: C, 79.31; H, 6.70; N, 6.17.

5,15-Bis(4,6-dimethoxypyrimidin-5-yl)-10,20-bis(2,4,6-trimethylphenyl)porphyrin (3b). To a well-stirred suspension of sodium hydride (80%, 0.040 g, 1.3 mmol) in DMF (5 mL) under argon was added methanol (0.06 mL, 1.48 mmol), and the mixture was stirred for 15 min at room temperature. A solution of porphyrin 3a (33.6 mg, 0.04 mmol) in DMF (5 mL) was added, and the mixture was stirred 60 °C for 40 h. The mixture was allowed to reach room temperature, water (30 mL) was added, and the mixture was extracted with dichloromethane (3 \times 30 mL). The combined organics were washed with water (3 imes 30 mL), dried over anhydrous magnesium sulfate, and evaporated under vacuum. The residue was chromatographed (silica gel/ dichloromethane) to afford porphyrin 3e which was recrystallized from dichloromethane/methanol (1:1) (0.015 g, 46%): ¹H NMR (400 MHz, CDCl₃) δ -2.51 (s, 2H), 1.85 (s, 12H), 2.62 (s, 6H); 3.82(s, 12H); 7.26(s, 4H), 8.64(d, J = 4.7 Hz, 4H), 8.67(s, 4H)2H), 8.91 (d, J = 4.7 Hz, 4H); UV-vis (CH₂Cl₂): $\lambda_{\text{max}} (\log \epsilon)$ 417.8 (5.6202), 514.0 (4.2505), 546.8 (3.7227), 590.2 (3.7282), 645.6 (3.4422). APCI-MS 883 (MH⁺). Anal. Calcd for C₅₀H₄₆N₈O₄: C, 72.97; H, 5.63; N, 13.62. Found: C, 72.85; H, 5.50; N, 13.60.

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